

**Effect of inoculation on the composting of source-selected organic
fraction of municipal solid wastes**

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Abstract

The effects of a commercial inoculum (MicroGest 10X,) on the field-scale composting of source-selected organic fraction of municipal solid wastes (OFMSW) have been studied by following routine parameters of the composting process (temperature, oxygen content and moisture) and biologically-related tests such as the respirometric index and the maturity grade. The inoculum was added to composting piles of OFMSW at different levels: Control (no added inoculum), Treatment A (10^5 CFU/g of OFMSW), Treatment B (10^6 CFU/g of OFMSW) and Treatment C (10^7 CFU/g of OFMSW).

The inoculum selected produced a significant acceleration of the composting process with high levels of biological activity in the thermophilic phase. In terms of composting acceleration and economical costs the optimal treatment was B, which produced a reduction of approximately half of the total composting time. Treatment C did not improve significantly the results obtained with treatment B, whereas treatment A has little effect on the OFMSW composting when compared to control experiment.

Respirometric index (determined at 55°C) and maturity grade appeared to be the most reliable tests to follow the biological activity of the OFMSW composting. On the other hand, routine parameters such as temperature, oxygen content and moisture showed no important differences among the applied treatments, and they should not be used in the determination of the effect of an inoculum in the composting process.

Keywords: Biological Activity, Composting, Stability, Inoculation, Maturity, Municipal Solid Wastes.

Introduction

In recent years, the international policy on management of organic wastes has been increasingly directed towards recycling. There are different technologies to recycle organic wastes from municipal solid wastes (mainly composed of kitchen wastes, pruning wastes and green wastes) and composting is often presented as a low-technology and low-investment process to convert organic wastes to an organic fertilizer known as compost.

Composting is a biotechnological process by which different microbial communities decompose organic matter into simpler nutrients. Composting is an aerobic process, which requires oxygen to stabilize the organic wastes, optimal moisture and porosity [1]. Temperature, oxygen and moisture are often selected as the control variables in the composting process, because they can be simply determined. However, there is scarce information about the monitoring of biological activity of composting processes in comparison with other biotechnology fields. Probably, the parameters most commonly used to describe the biological activity of a composting process or a compost are stability and maturity. Stability is related to the presence of easily biodegradable compounds and is routinely measured by the respirometric index as the Oxygen Uptake Rate (OUR) of a compost sample [2,3]. Maturity is usually determined with self-heating tests in Dewar vessels [4] or germination indexes [5], and it is used in relation to compost application and plant growth [6].

As a biotechnological process, composting needs inoculation. Nevertheless, most of the traditionally composted wastes, such as source-selected OFMSW, sewage sludge or animal manure, are supposed to contain enough autochthonous microorganisms and thus inoculation is rarely used in composting. In fact, opposite results on the inoculation of different composting processes can be found in literature. In general, from previous studies,

it seems that when a specialized inoculum is used some positive effects can be observed. For instance, it has been reported that a bacterial inoculum enriched with feather-degrading bacteria enhances keratin degradation and biofilm formation in poultry compost [7], or that the biodegradability of poly-caprolactone in a compost environment is significantly altered by the type of inoculum used [8]. However, other similar works have not found significant improvements when some inoculum was used in the composting of different wastes. The failure of inoculum in these works is attributed to different reasons, such as the presence of sufficient quantity of microorganisms in composted materials for an adequate composting [9,10] or competition between autochthonous and inoculated strains [11]. In other works, although there is no perceptible effect of inoculation, it is pointed that inoculation decreases the variability observed in the composting experiences [12].

Nevertheless in most of the cited works only routine parameters of the composting process are profiled and compared between inoculated and non-inoculated treatments. Moreover, only an inoculum dosage is usually tested. In the few works when microbiological parameters are determined, it seems clear that inoculation can have a positive effect in the composting acceleration and the quality of the compost, especially in the first thermophilic stage of the process [13,14].

The objective of this work is to study systematically the effect of different dosages of a commercial inoculum (MicroGest 10X,) in the acceleration of the composting process of OFMSW. The study is focused on the biological activity of the OFMSW composting (measured as respirometric index and maturity grade), since this is the key parameter for the determination of an inoculum performance.

Materials and Methods

Organic wastes

Source-selected organic fraction of municipal solid wastes (OFMSW) was used as main substrate for composting experiments. OFMSW was obtained and composted at the composting plant of Jorba (Barcelona, Spain). Plastic, metal, glass and other visible contaminants of the organic fraction were manually removed.

Inoculum composition and preparation

A commercial inoculum named MicroGest 10X was generously provided by MicroGest 10X is a combination of enzymes and microorganisms obtained from dried *Bacillus* fermentation product, dried *Saccharomyces cerevisiae* fermentation product, dried *Trichoderma reesei* extract, dried *Aspergillus niger* extract, dried whey and silicon dioxide. The minimum number of total *Bacillus* bacteria is $4 \cdot 10^9$ CFU/g of MicroGest 10X.

MicroGest 10X was blended with distilled water (room temperature) and added to the solid waste by direct irrigation on the material received at the composting plant. All the composting material was turned after inoculation to spread the bacterial consortium. The treatments applied to the OFMSW piles were: Control (no added inoculum), A (10^5 CFU/g of OFMSW), B (10^6 CFU/g of OFMSW) and C (10^7 CFU/g of OFMSW).

Composting experiments

Piles of source-separated OFMSW were investigated. The piles were built according to the normal operation of the plant. Trapezoidal piles (base: 2 m; height: 1.5 m) of approximately 30 t of OFMSW mixed with pruning wastes in a 2:1 volumetric ratio were

used for each treatment of inoculum applied. Piles were turned weekly using a turner Backhus Model 15.50. No forced aeration was provided.

Pile temperature was measured at 50 and 100 cm depth and pile oxygen content was measured at 100 and 150 cm depth in 4 points of the pile. Temperature and oxygen values are presented as average values. Variability between temperature and oxygen values measured at different points of the pile was in the range of 10-20% (error bars not included in figures due to space limitations). Temperature was measured with a portable Pt-100 sensor (Delta Ohm HD9214) and oxygen concentration in interstitial air was measured with a portable O₂ detector (Oxy-ToxiRAE, RAE) connected to a portable aspiration pump. Other parameters of the composting process were analyzed in the laboratory after extracting a representative solid sample (approximately 2-L volume) of the pile from 4 different points.

Respirometric tests

A static respirometer was built according to an original model previously described [2] and following the modifications and recommendations given by the U.S. Department of Agriculture and U.S. Composting Council [15].

Approximately 250 mL of compost samples (representative sample of the pile from 4 different locations) were placed in 500 mL Erlenmeyer flasks on a nylon mesh screen that allowed air movement under and through the solid samples. The setup included a water bath to maintain the selected temperature during the respirometric test. In the experiments presented, respirometric tests were obtained at two temperatures: 55°C (related to the actual biological activity of the process) and 37°C (related to the stability of the material). Prior to

the assays, samples were incubated for 18 hours at 37°C or 4 hours at 55°C. During all the incubation period samples were aerated with previously humidified air at the sample temperature. The drop of oxygen content in a flask containing a compost sample was monitored with a dissolved oxygen meter (Lutron 5510, Lutron Co. Ltd., Taiwan) connected to a data logger. The rate of respiration of the compost sample (Oxygen Uptake Rate, OUR, based on total organic matter content, TOM) was then calculated from the slope of oxygen level decrease according to the standard procedures [15]. Results of the static respirometric index referred to total organic matter content are presented as an average of three replicates. The standard deviation between respirometric replications is in the range of 5-10% (error bars not included in figures due to space limitations).

Analytical Methods

Moisture, total organic matter (TOM), pH, electrical conductivity (EC), Solvita® test and Rottegrade self-heating test were determined according to the standard procedures [15]. Representative samples of the pile from 4 different locations were used to carry out all the analytical tests.

Data analysis

Variance analysis was done to compare mean values of different parameters studied for each treatment applied using the least-significance difference test at 5% level of probability.

Results and Discussion

Evolution of composting experiments

Different routine parameters were determined in the composting experiments. Figure 1 presents the temperature profiles obtained for the different experiments (average values). Thermophilic range of temperatures ($>50^{\circ}\text{C}$) could be easily reached for all the dosages of inoculum, including the control experiment. According to Figure 1, temperature-time profiles indicated that the material should be hygienized. However, no significant differences ($P\leq 0.05$) were observed among the treatments used.

Figure 2 shows the evolution of interstitial oxygen content of the material for each treatment. Although the oxygen profiles were similar and differences among average oxygen values were not statistically significant ($P\leq 0.05$), it seemed that high dosages of inoculum implied lower oxygen concentrations in some samples, which was in accordance with higher microbial aerobic activity. Nevertheless, all experiments showed an oxygen profile very similar to those found in typical windrow composting [1], with oxygen limitation in the first stage (some oxygen values below 5%) and a final stage with high concentrations of oxygen (corresponding to a decrease in biological activity).

On the other hand, moisture quickly decreased for all the dosages (data not shown) according to the high temperatures reached, and water addition was necessary after 20-30 days of composting to maintain a moisture level over 40%. No significant differences were detected for each treatment ($P\leq 0.05$). In conclusion, it can be stated that there were no significant differences among the treatments using different doses of inoculum in relation to the common composting parameters, such as temperature, oxygen content and moisture.

The profiles obtained for these parameters were very similar to those found in typical windrow composting of OFMSW.

Physico-chemical characteristics

Moisture, TOM, EC and pH were determined for the initial sample of OFMSW and the final compost obtained for each treatment (after 100 days of composting). Results obtained are presented in Table 1. As can be seen in Table I, only slight differences were observed among the treatments applied. In fact, this variability is usually observed in different batches of standard non-inoculated batches of OFMSW composting [12,16]. Therefore, it could be concluded that inoculation did not have a significant impact on these parameters.

Biological activity indexes

Different measures of biological activity related to maturity and stability were used to follow the material evolution and include respirometric index (measured at 37°C and 55°C), Rottegrade self-heating test and Solvita® test. These tests have been extensively used in the composting field to characterize the stability and maturity of composts [2,4,17]. However, the information they provide are qualitatively different. Thus, respirometry refers to the aerobic biological activity of the material, and it is recommended to monitor the composting process [18,19]. Also, it can be determined at two different temperatures: 55°C (which is a measure of the real activity of the process in the thermophilic range) or 37°C (which is related to the stability of the material). It is generally considered that values of respirometric index (measured at 37°C) below $1 \text{ mg O}_2 \cdot \text{g TOM}^{-1} \cdot \text{h}^{-1}$ correspond to stable

compost [20]. On the other hand, Rottegrade self-heating test and Solvita tests give information in form of maturity grade. In Europe, the Rottegrade self-heating test is commonly used to quantify the maturation of compost, with a range from I (fresh material) to V (compost completely mature) [15]. The results of the respirometric index (at 37°C and 55°C) and maturity grade obtained for each treatment are presented in Figures 3, 4 and 5, respectively.

From Figures 3 and 4, it was obvious that doses B and C of inoculum produced a faster decrease of the respirometric index (measured at 37°C and 55°C) than those observed for dose A and control, indicating a clear acceleration of the whole process. This fact can be due to a high microbial activity in the composting thermophilic phase, which finally produced a more stable product. As expected, it was also clear that between doses B and C, dose C is more effective. It can be concluded that larger quantities of inoculum accelerates the composting of OFMSW, however, the differences in the respirometric index between doses B and C probably are not high enough from the economical point of view. Finally, dose A does not seem to have a significant effect on the respirometric index.

As expected, when respirometric indexes were determined at 37°C and 55°C, differences between both indexes were more significant in the first thermophilic phase than in the final maturation phase, when the temperature was close to 37°C. In fact, respirometric indexes determined at 37°C were not statistically different ($P \leq 0.05$) from those determined at 55°C from day 50 on (corresponding to mesophilic phase, Figure 1). It seemed evident that the thermophilic microorganisms only exhibited a limited growth at 37°C, whereas the mesophilic population was scarce. At 55°C, the respirometric index was determined under thermophilic conditions and the microbial populations present in the material were fully

active, resulting in high values of the respirometric index. In conclusion, it can be stated that the respirometric index can be used for monitoring the biological activity of the composting process; however, it should be determined at thermophilic conditions, whereas determinations at mesophilic temperature should be exclusively used for compost material in the maturation stage.

On the other hand, results from Figure 5 (maturity grade) are very interesting because they confirm the results obtained for the respirometric index. Thus, it took 8 weeks for the control experiment to reach a maturity grade considered stable (IV), which is a typical time in the windrow composting of OFMSW. In the inoculated experiments, the acceleration in the process to obtain mature compost was spectacular. With dose B, it only took 4 weeks to obtain grade IV compost, and 7 weeks to obtain grade V (the maximum maturity grade). With dose A, the results were less positive, but better than control experiment. The results for dose C were similar to those obtained in experiment B, which indicated that the optimal dose in terms of acceleration and cost was dose B. The optimal treatment in terms of maturity grade is therefore dose B, which produced a reduction of the 50% of the composting time.

Finally, Solvita test was studied for initial and final samples of compost (Table 1). This test is based on the qualitative determinations of CO₂ and NH₃ emissions from a compost sample, which are correlated to a maturity grade (one for CO₂ and one for NH₃, ranging from 0 to 8). From our results, there were no differences among the treatments using the Solvita test, and all the materials appeared to be only partially stabilized. These results were contradictory with respirometric index and maturity grade. It is probable that

this test is not suitable to assess the stability of heterogeneous materials such as OFMSW, although it has been successfully used with other wastes [17].

Conclusions

From the results obtained, it can be concluded that:

- 1) MicroGest 10X is an effective inoculum to accelerate the composting of OFMSW, by enhancing the biological activity in the thermophilic stage of the process.
- 2) Among the inoculum dosages tested, treatment B (10^6 CFU/g of OFMSW) is the optimal in terms of reduction of the composting time (approximately 50% reduction) and economics of the treatment. Treatment C (10-fold more inoculum amount than treatment B) does not significantly improve the composting of OFMSW in comparison to treatment B, whereas treatment A (10-fold less inoculum amount than treatment B) has little effect when compared to control experiment. The determination of an optimal dosage of inoculum appears to be as important as the decision to use it.
- 3) Routine parameters (temperature, oxygen content and moisture) are not statistically different among the treatments studied; therefore, they should not be used in evaluating the performance of an inoculum in the composting of OFMSW.
- 4) Parameters related with biological activity of the material such as stability and maturity are specially indicated to study the effects of inoculation of composting materials. Concretely, respirometric index (determined at 55°C) and maturity grade can be considered as reference parameters in the composting field.

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References

1. Haug RT. *The Practical Handbook of Compost Engineering*. Boca Raton, FL: Lewis Publishers, 1993.
2. Iannotti DA, Pang T, Toth BL, Elwell DL, Keener HM, Hoitink HAJ. A quantitative respirometric method for monitoring compost stability. *Compost Sci Util* 1993;1:52-65.
3. Chica A, Mohedo JJ, Martín MA, Martín A. Determination of the stability of MSW compost using a respirometric technique. *Compost Sci Util* 2003;11:169-175.
4. Weppen P. Determining compost maturity: evaluation of analytical properties. *Compost Sci Util* 2002;10:6-15.
5. Bernal MP, Paredes C, Sánchez-Monedero MA, Cegarra J. Maturity and stability parameters of composts prepared with a wide range of organic wastes. *Bioresource Technol* 1998;63:91-99.
6. Cooperband LR, Stone AG, Fryda MR, Ravet JL. Relating compost measures of stability and maturity to plant growth. *Compost Sci Util* 2003;11:113-124.
7. Ichida JM, Krizova L, LeFevre CA, Keener HM, Elwell DL, Burt EH. Bacterial inoculum enhances keratin degradation and biofilm formation in poultry compost. *J Microbiol Meth* 2001;47:199-208.

8. Ohtaki A, Akakura N, Nakasaki, K. Effects of temperature and inoculum on the degradability of poly-caprolactone during composting. *Polym Degrad Stabil* 1998;62:279-284.
9. Faure D, Deschamps AM. The effect of bacterial inoculation on the initiation of composting of grape pulps. *Bioresource Technol* 1991;37:235-238.
10. Nakasaki K, Watanabe A, Kitano M, Kubota H. Effect of seeding on thermophilic composting of tofu refuse. *J Environ Qual* 1992;21:714-719.
11. Baheri H, Meysami P. Feasibility of fungi bioaugmentation in composting a flare pit soil. *J Hazard Mater* 2002;B89:279-286.
12. Schloss PD, Walker LP. Measurement of process performance and variability in inoculated composting reactors using ANOVA and power analysis. *Process Biochem* 2000;35:931-942.
13. Tiquia SM, Tam NFY, Hodgkiss IJ. Effects of bacterial inoculum and moisture adjustment on composting pig manure. *Environ Pollut* 1997;96:161-171.
14. Bolta SV, Mihelic R, Lobnik F, Lestan D. Microbial community structure during composting with and without mass inocula. *Compost Sci Util* 2003;11:6-15.
15. U.S. Department of Agriculture and U.S. Composting Council. Test methods for the examination of composting and compost. Houston, TX: Edaphos International, 2001.
16. Schloss PD, Chaves B, Walker LP. The use of the analysis of variance to assess the influence of mixing during composting. *Process Biochem* 2000;35:675-684.

17. Changa CM, Wang P, Watson ME, Hoitink HAJ, Michel FC. Assessment of the reliability of a commercial maturity test kit for composted manures. *Compost Sci Util* 2003;11:125-143.
18. Mari I, Ehaliotis C, Kotsou M, Balis C, Georgakakis D. Respiration profiles in monitoring the composting of by-products from the olive oil agro-industry. *Bioresource Technol* 2003;87:331-336.
19. Liang C, Das KC, McClendon RW. The influence of temperature and moisture contents regimes on the aerobic microbial activity of a biosolids composting blend. *Bioresource Technol* 2003;86:131-137.
20. California Compost Quality Council. CCQC-Compost Maturity Index. Available: <http://www.ccqc.org/Documents/MatIndex.pdf> [2002, October 10].

Tables

Table 1: Initial characteristics of OFMSW composted and final characteristics of the compost obtained for each treatment studied.

Parameter	Initial	Control	Treatment A	Treatment B	Treatment C
Moisture (%)	57.98	35.5	35.2	33.1	24.2
TOM (%)	62.86	45.8	47.7	44.4	46.8
pH	6.90	8.60	8.33	8.48	8.90
EC (mS/cm)	2.97	5.25	4.92	5.57	4.82
Solvita CO ₂	3	5	5	5	5
Solvita NH ₃	4	5	4	4	4

Figure Legends

Figure 1: Temperature profiles (average values) for the treatments studied. Circles: Control experiment; Diamond: Treatment A; Square: Treatment B; Triangle: Treatment C.

Figure 2: Oxygen content profiles (average values) for the treatments studied. Circles: Control experiment; Triangle: Treatment A; Square: Treatment B; Diamond: Treatment C.

Figure 3: Respirometric index (37°C) profiles for the treatments studied. Circles: Control experiment; Triangle: Treatment A; Square: Treatment B; Diamond: Treatment C.

Figure 4: Respirometric index (55°C) profiles for the treatments studied. Circles: Control experiment; Triangle: Treatment A; Square: Treatment B; Diamond: Treatment C.

Figure 5: Maturity grade profiles for the treatments studied. Circles: Control experiment; Triangle: Treatment A; Square: Treatment B; Diamond: Treatment C.

Figure 1: Barrena et al.

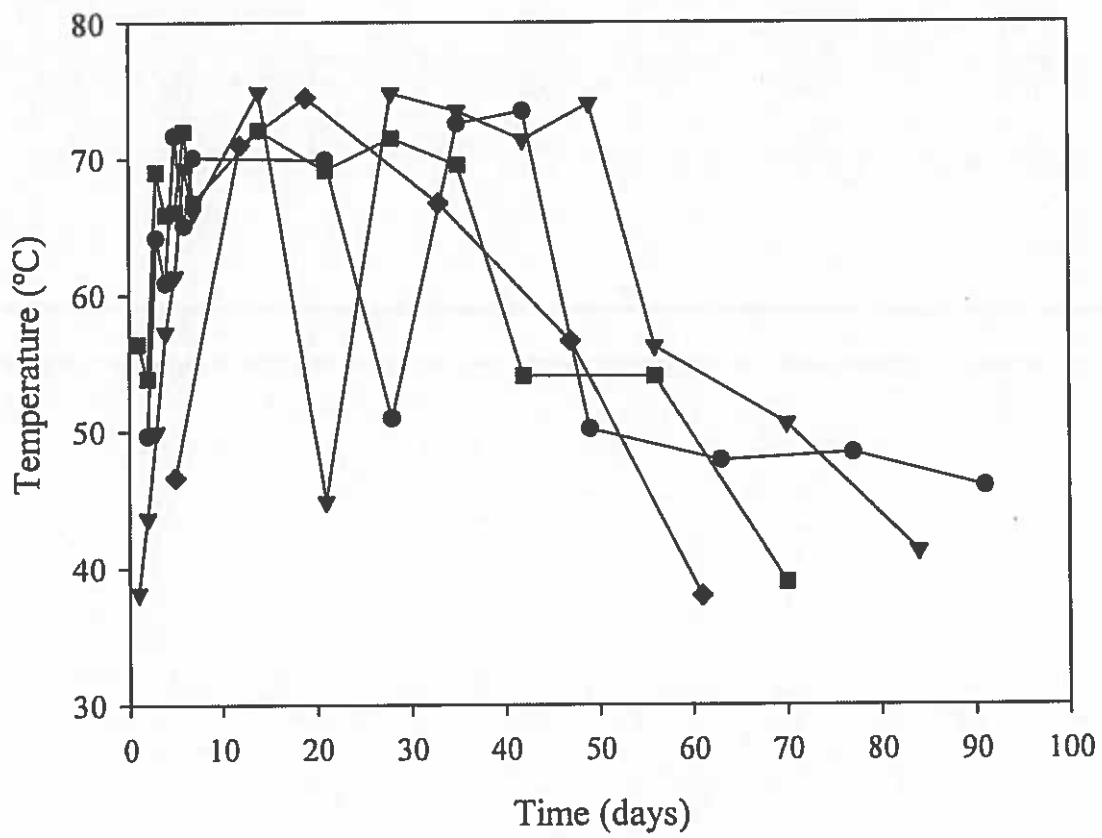


Figure 2: Barrena et al.

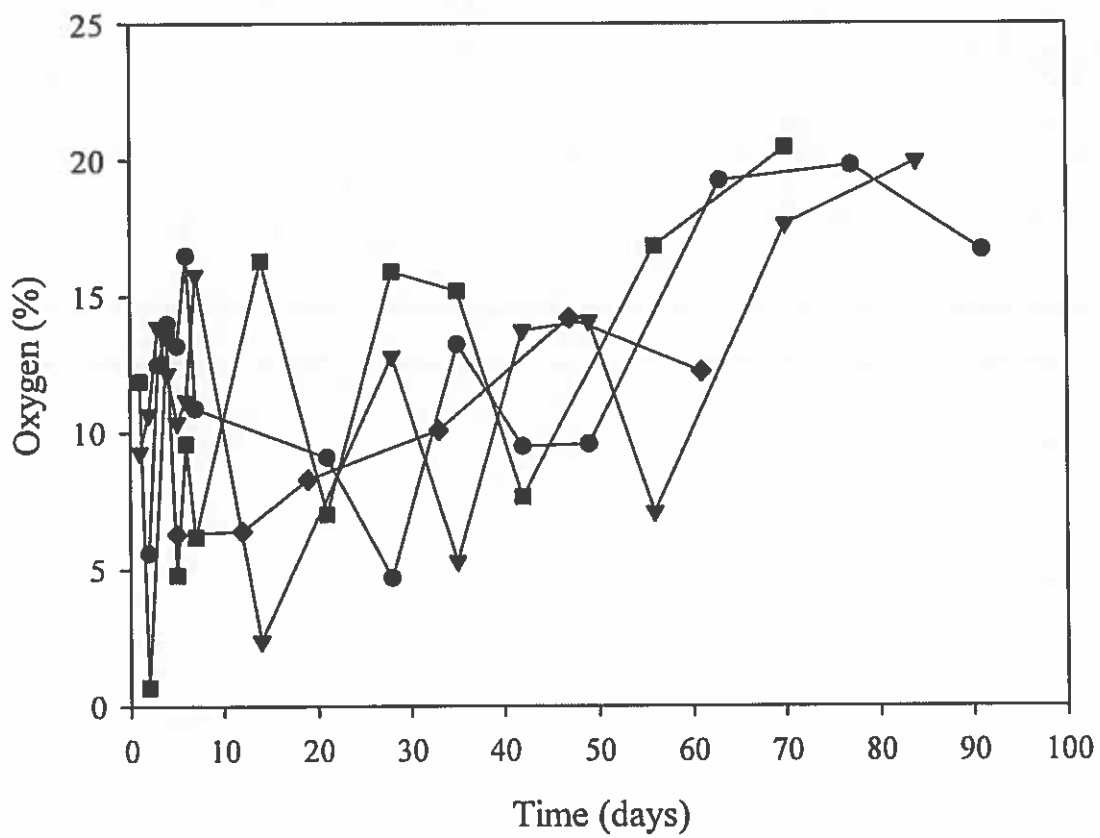


Figure 3: Barrena et al.

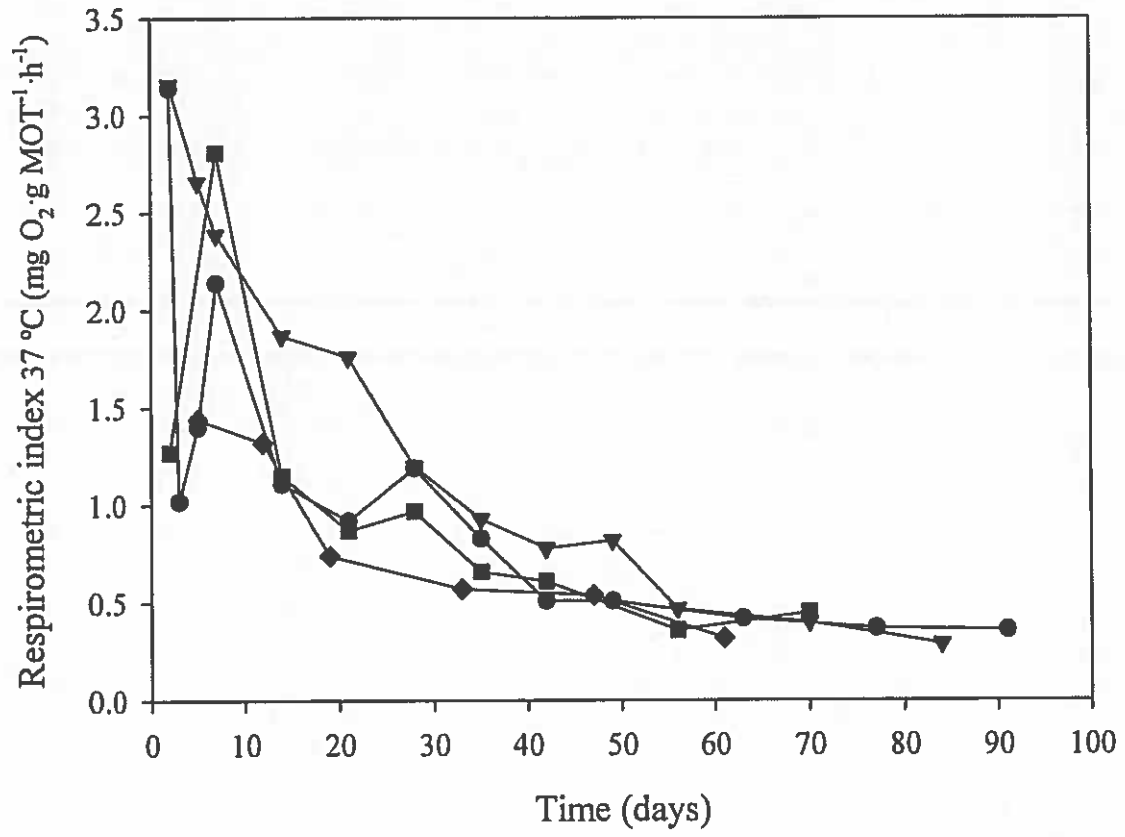


Figure 4: Barrena et al.

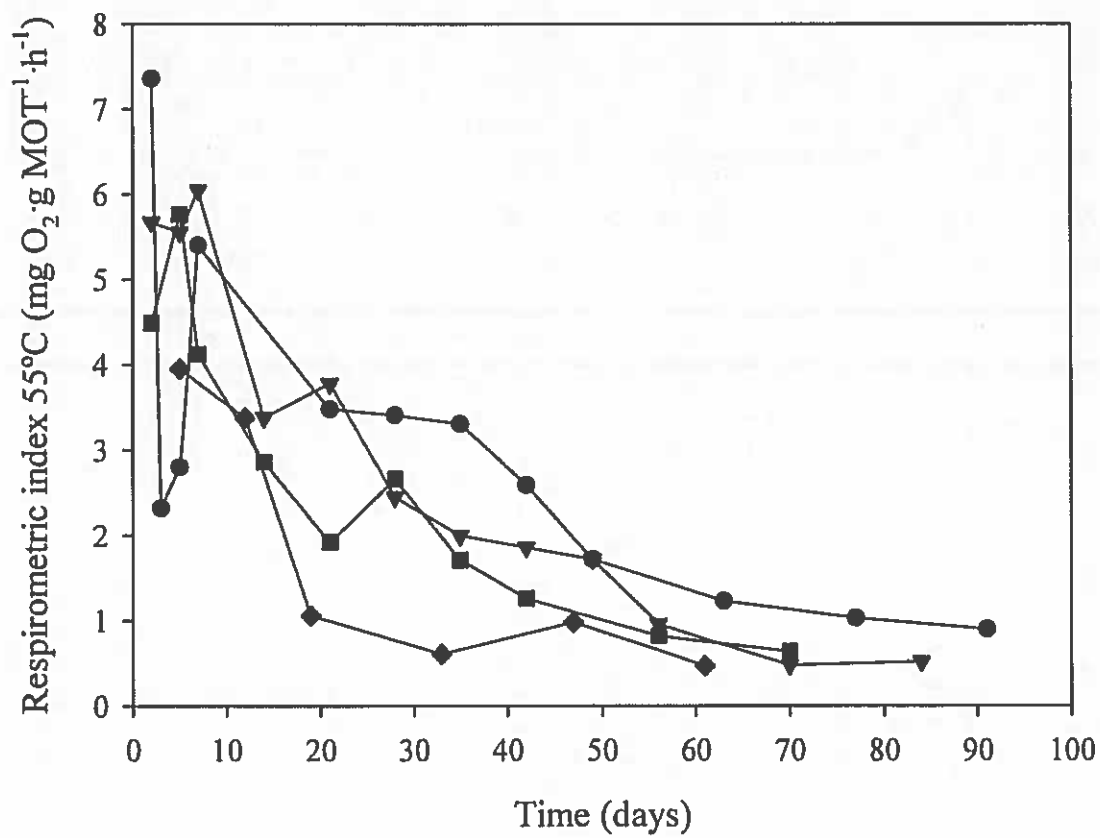


Figure 5: Barrena et al.

